# Woods Hole 2015 - brief primer on Multiple Sequence Alignment – Mark Holder

Many forms of sequence alignment are used in bioinformatics:

- Structural Alignment

- Local alignments

- **Global, evolutionary alignment**
  - Inputs: unaligned sequences thought to be homologous over their full length
  - we often ignore events like transpositions or inversions

The goal of MSA is to introduce gaps such that:

- residues in the same column are homologous – all descended from a residue in the common ancestor, and
- all descendants of a residue are put in the same column.

slide by Derrick Zwickl

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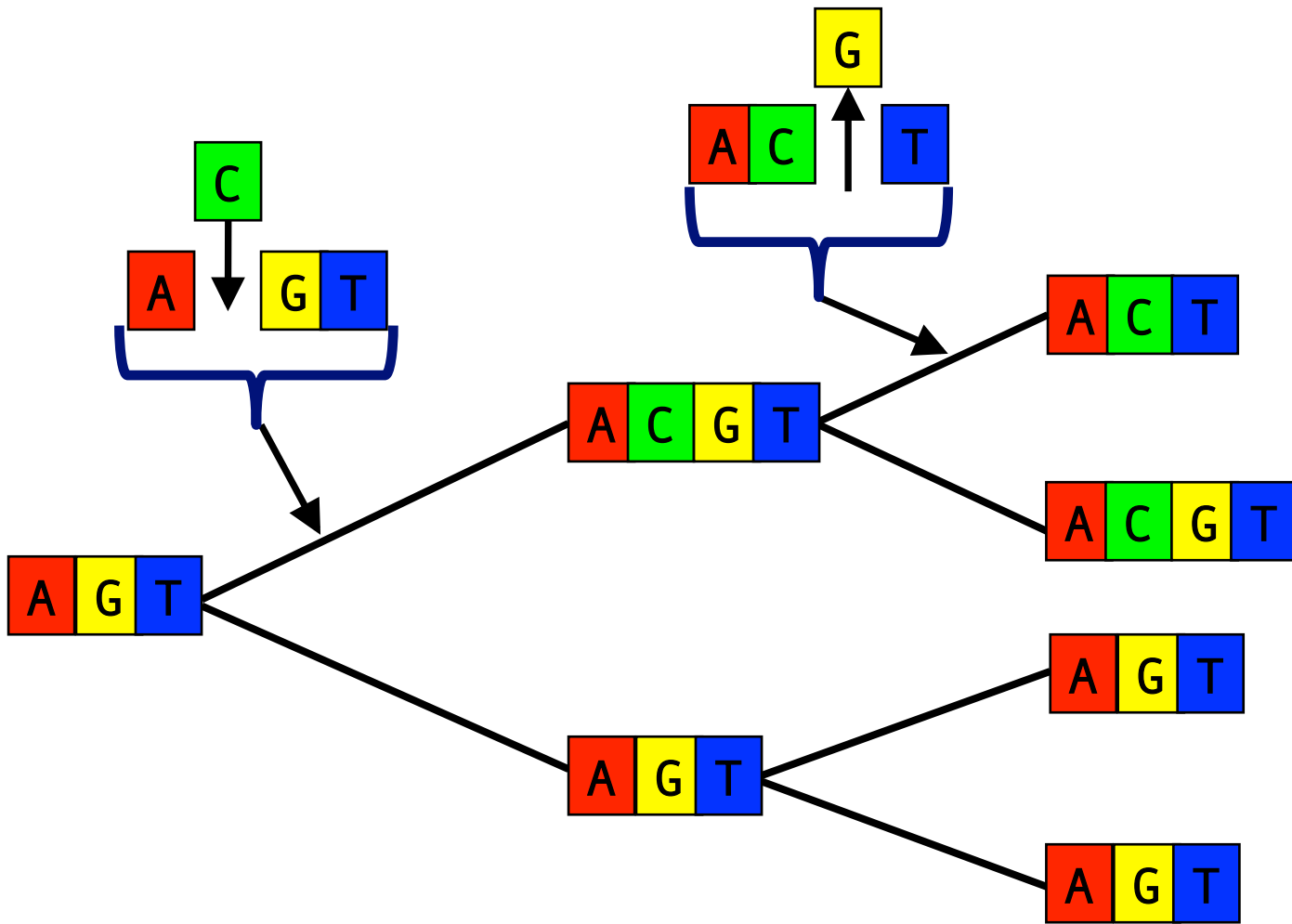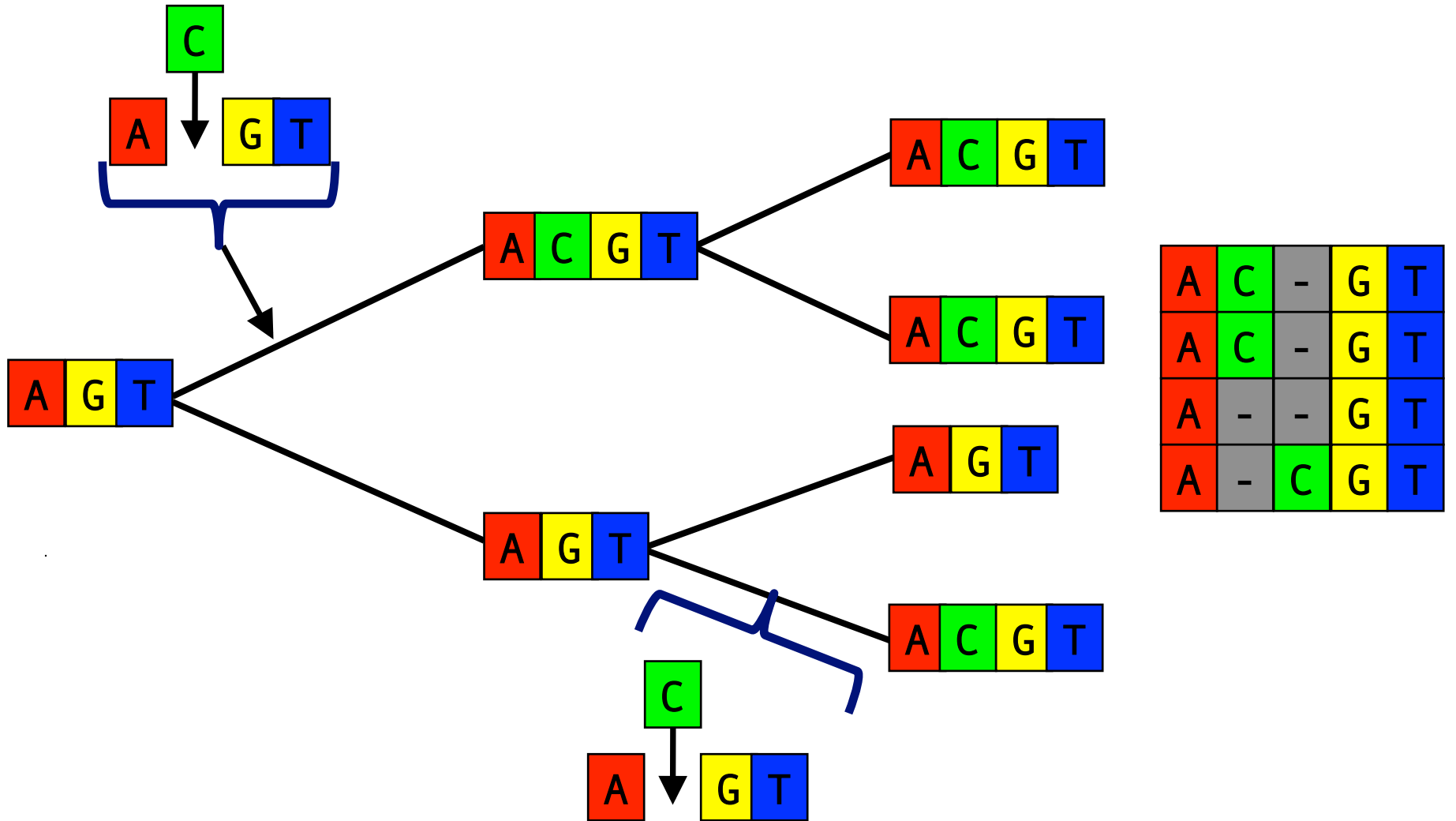
Main points

- Accuracy of alignment is important.
- Pairwise alignment is tractable.
- Most MSA programs use progressive alignment:
  - a series of pairwise operations.
  - these algorithms are **not** guaranteed to return the optimal solution.
  - the criteria used are not ideal from an evolutionary standpoint.

Main points (continued)

- Simultaneous inference of MSA and tree is the most appropriate choice, but is computationally demanding. See: Poisson Indel Process (Bouchard-Côté and Jordan, 2013), Bali-Phy, Handel, AliFritz, and POY software
- Many people cull ambiguously aligned regions.

```
human    KRSV
chimp    KRV
gorilla  KSV
orang    KPRV
```

How we align these sequences affects tree estimation.

human KRSV		human KRSV		human KRSV
chimp KR-V		chimp K-RV		chimp KR-V
gorilla KS-V		gorilla K-SV		gorilla K-SV
orang KPRV		orang KPRV		orang KPRV

Pairwise alignment

Gap penalties and a substitution matrix imply a score for any alignment. Pairwise alignment involves finding the alignment that maximizes this score.

- substitution matrices assign positive values to matches or substitutions between similar residues (for example Leucine→Isoleucine).
- infrequent types of substitutions receive negative scores
- indels are rare, so gaps are heavily penalized (negative scores).

BLOSUM 62 Substitution matrix

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
A	4																			
R	-1	5																		
N	-2	0	6																	
D	-2	-2	1	6																
C	0	-3	-3	-3	9															
Q	-1	1	0	0	-3	5														
E	-1	0	0	2	-4	2	5													
G	0	-2	0	-1	-3	-2	-2	6												
H	-2	0	1	-1	-3	0	0	-2	8											
I	-1	-3	-3	-3	-1	-3	-3	-4	-3	4										
L	-1	-2	-3	-4	-1	-2	-3	-4	-3	2	4									
K	-1	2	0	-1	-3	1	1	-2	-1	-3	-2	5								
M	-1	-1	-2	-3	-1	0	-2	-3	-2	1	2	-1	5							
F	-2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6						
P	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7					
S	1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4				
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	1	5			
W	-3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	1	-4	-3	-2	11		
Y	-2	-2	-2	-3	-2	-1	-2	-3	2	-1	-1	-2	-1	3	-3	-2	-2	2	7	
V	0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	-2	0	-3	-1	4
	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V

Scoring an alignment with the BLOSUM 62 matrix

Pongo	V	D	E	V	G	G	E	L	G	R	L	F	V	V	P	T	Q
Gorilla	V	E	V	A	G	D	L	G	R	L	L	I	V	Y	P	S	R
Score	4	2	-2	0	6	-6	-3	-4	-2	-2	4	0	4	-1	7	4	1

The score for the alignment is

$$D_{ij} = \sum_k d_{ij}^{(k)}$$

If i indicates *Pongo* and j indicates *Gorilla*. (k) is just an index for the column.

$$D_{ij} = 12$$

Scoring an alignment with gaps

If we were to use a gap penalty of -8:

Pongo	V	D	E	V	G	G	E	L	G	R	L	–	F	V	V	P	T	Q
Gorilla	V	–	E	V	A	G	D	L	G	R	L	L	I	V	Y	P	S	R
Score	4	–8	5	5	0	6	2	4	6	5	4	–8	0	4	–1	7	4	1

By introducing gaps we have improved the score:

$$D_{ij} = 40$$

Gap Penalties

Penalizing gaps more heavily than substitutions avoids alignments like this:

```
Pongo    VDEVGGE-LGRLFVVPTQ
Gorilla  VDEVGG-DLGRLFVVPTQ
```
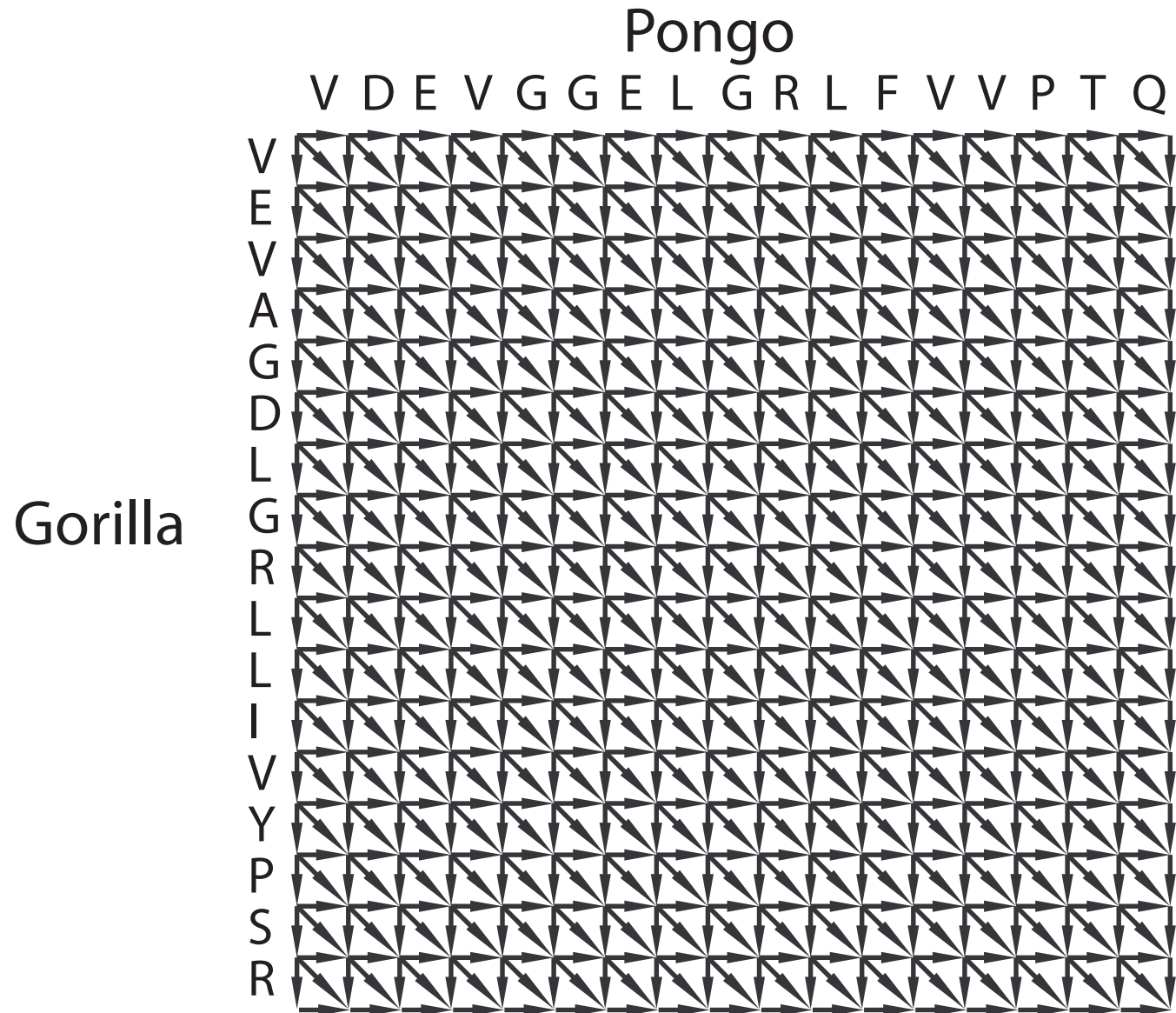
Affine gap penalties are often used to accommodate multi-site indels:

$$GP = GO + (l)GE$$

where:

- GP is the gap penalty.

- GO is the "gap-opening penalty"

- GE is the "gap-extension penalty"

- l is the length of the gap
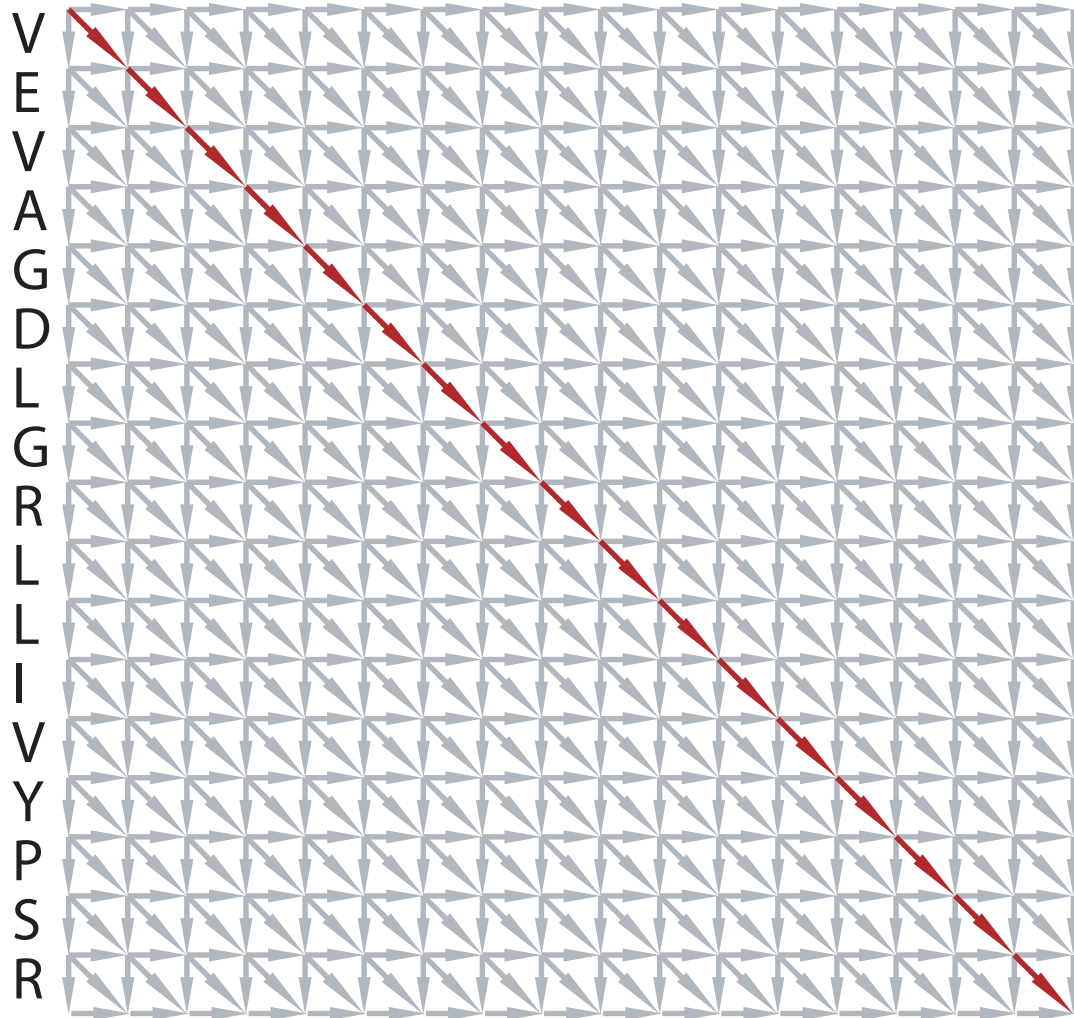
Finding an optimal alignment

	pongo	V	D	E	V	G	G	E	L	G	R	L	F	V	V	P	T	Q
	gorilla	V	E	V	A	G	D	L	G	R	L	L	I	V	Y	P	S	R
	score	4	2	-2	0	6	-6	-3	-4	-2	-2	4	0	4	-1	7	4	1

	Pongo	V	D	E	V	G	G	E	L	G	R	L	–	F	V	V	P	T	Q
	Gorilla	V	–	E	V	A	G	D	L	G	R	L	L	I	V	Y	P	S	R
	Score	4	-8	5	5	0	6	2	4	6	5	4	-8	0	4	-1	7	4	1

Needleman-Wunsch algorithm (paraphrased)

- Work from the top left (beginning of both sequences)
- For each cell store the highest score possible for that cell and a "back" pointer to tell point to the previous step in the best path
- When you reach the lower right corner, you know the optimal score and the back pointers tell you the alignment.
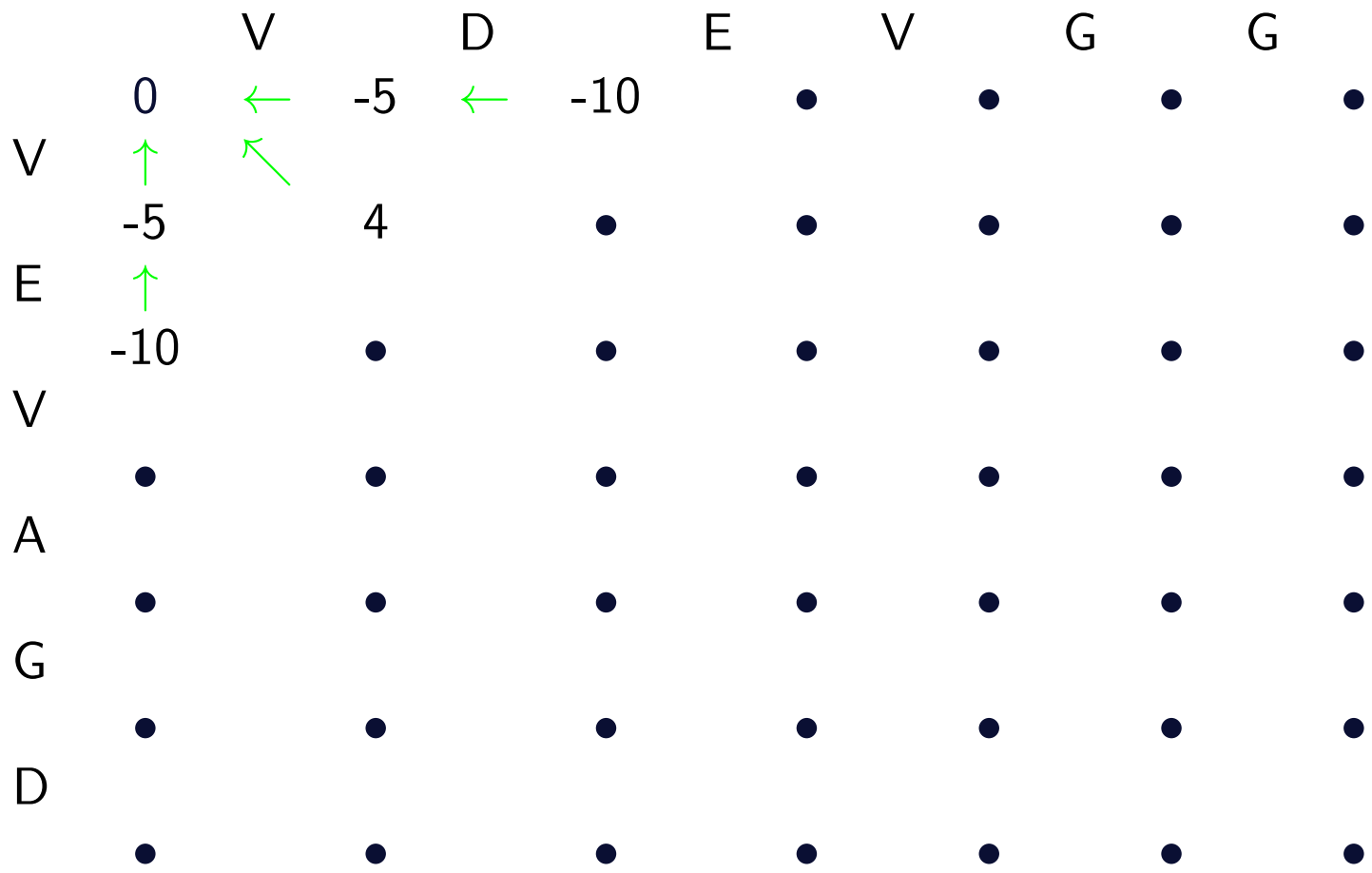
The highest score calculation at each cell only depends on the cell's 3 possible previous neighbors.

If one sequence is length M_1, and the other is length M_2, then Needleman-Wunsch only takes $\mathcal{O}(M_1 M_2)$ calculations.

But there are a **much** larger number of possible alignments (# alignments grows exponentially).

		V	D	E	V	G	G
	0	•	•	•	•	•	•
V	•	•	•	•	•	•	•
E	•	•	•	•	•	•	•
V	•	•	•	•	•	•	•
A	•	•	•	•	•	•	•
G	•	•	•	•	•	•	•
D	•	•	•	•	•	•	•

Using a gap penalty of -5

	V	D	E	V	G	G
0 ← -5	•	•	•	•	•
V ↑ -5	•	•	•	•	•	•
E •	•	•	•	•	•	•
V •	•	•	•	•	•	•
A •	•	•	•	•	•	•
G •	•	•	•	•	•	•
D •	•	•	•	•	•	•

		V	D	E	V	G	G
	0	← -5	← -10	•	•	•	•
V	↑ ↖						
	-5	4	•	•	•	•	•
E	↑						
	-10	•	•	•	•	•	•
V							
	•	•	•	•	•	•	•
A							
	•	•	•	•	•	•	•
G							
	•	•	•	•	•	•	•
D							
	•	•	•	•	•	•	•

		V	D	E	V	G	G
	0 ←	-5 ←	-10 ←	-15	•	•	•
V	↑	↖					
	-5	4 ←	-1	•	•	•	•
E	↑	↑					
	-10	-1	•	•	•	•	•
V	↑						
	-15	•	•	•	•	•	•
A							
	•	•	•	•	•	•	•
G							
	•	•	•	•	•	•	•
D							
	•	•	•	•	•	•	•

		V	D	E	V	G	G
	0 ←	-5 ←	-10 ←	-15 ←	-20	●	●
V	↑	↖					
	-5	4 ←	-1 ←	-6	●	●	●
E	↑	↑	↖				
	-10	-1	6	●	●	●	●
V	↑	↖	↑				
	-15	-6	●	●	●	●	●
A	↑						
	-20	●	●	●	●	●	●
G							
	●	●	●	●	●	●	●
D							
	●	●	●	●	●	●	●

Pairwise alignment is a beautiful topic in bioinformatics

- Clever programming tricks let us find the best-scoring pairwise alignment quickly

- The additive scoring system:

  - can incorporate biological knowledge (via empirically-based substitution matrices)
  - can almost be justified in terms of powerful statistical methodology (maximum likelihood).

Pairwise alignment costs

- Paul Lewis will explain likelihood tomorrow,
- Additive costs can be justified as approximations to the log of likelihoods if:
  - we can identify the events that must have occurred in generate the data, and
  - we can assign (relative) probabilities based on whether these events are rare or common.

Pongo	V	D	E	V	G	G	E	L	G	R	L	-	F	V	V	P	T	Q
Gorilla	V	-	E	V	A	G	D	L	G	R	L	L	I	V	Y	P	S	R
Score	4	-8	5	5	0	6	2	4	6	5	4	-8	0	4	-1	7	4	1
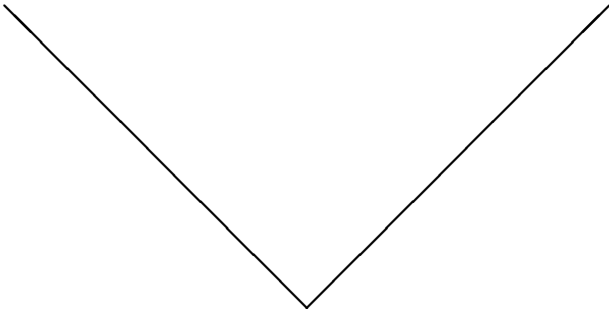
Pongo   *Gorilla*

Pongo	V	D	E	V	G	G	E	L	G	R	L	–	F	V	V	P	T	Q
Gorilla	V	–	E	V	A	G	D	L	G	R	L	L	I	V	Y	P	S	R
Score	4	-8	5	5	0	6	2	4	6	5	4	-8	0	4	-1	7	4	1
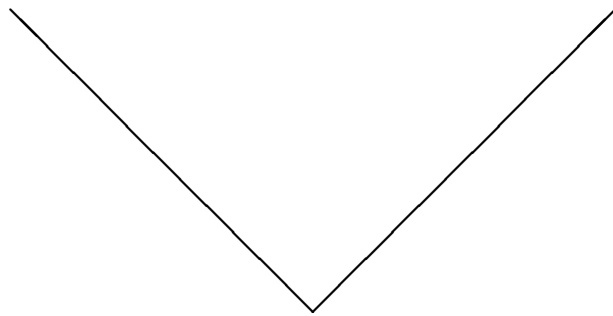
Pongo   *Gorilla*

$$V \leftrightarrow V \qquad \mathbb{P}(\text{pos. } 1) = \mathbb{P}(V \leftrightarrow V)$$

Pongo	**V**	**D**	E	V	G	G	E	L	G	R	L	–	F	V	V	P	T	Q
Gorilla	**V**	-	E	V	A	G	D	L	G	R	L	L	I	V	Y	P	S	R
Score	4	-8	5	5	0	6	2	4	6	5	4	-8	0	4	-1	7	4	1

Pongo   *Gorilla*

V $\leftrightarrow$ V
D $\leftrightarrow$ -

$$\mathbb{P}(\text{pos. } 1-2) = \mathbb{P}(V \leftrightarrow V)$$
$$\times \mathbb{P}(D \leftrightarrow -)$$

	V	D	E	V	G	G	E	L	G	R	L	–	F	V	V	P	T	Q
Pongo	**V**	**D**	**E**	V	G	G	E	L	G	R	L	–	F	V	V	P	T	Q
Gorilla	**V**	-	**E**	V	A	G	D	L	G	R	L	L	I	V	Y	P	S	R
Score	4	-8	5	5	0	6	2	4	6	5	4	-8	0	4	-1	7	4	1

Pongo   *Gorilla*

$$
\begin{aligned}
\text{V} &\leftrightarrow \text{V}\\
\text{D} &\leftrightarrow \text{–}\\
\text{E} &\leftrightarrow \text{E}
\end{aligned}
$$

$$
\begin{aligned}
\mathbb{P}(\text{pos. } 1-3) &= \mathbb{P}(V \leftrightarrow V)\\
&\times \mathbb{P}(D \leftrightarrow -)\\
&\times \mathbb{P}(E \leftrightarrow E)
\end{aligned}
$$

	V	D	E	V	G	G	E	L	G	R	L	–	F	V	V	P	T	Q
Pongo	**V**	**D**	**E**	V	G	G	E	L	G	R	L	–	F	V	V	P	T	Q
Gorilla	**V**	-	**E**	V	A	G	D	L	G	R	L	L	I	V	Y	P	S	R
Score	4	-8	5	5	0	6	2	4	6	5	4	-8	0	4	-1	7	4	1

Pongo   *Gorilla*

$$
\begin{aligned}
\text{V} &\leftrightarrow \text{V} \\
\text{D} &\leftrightarrow \text{-} \\
\text{E} &\leftrightarrow \text{E}
\end{aligned}
$$

$$
\begin{aligned}
\ln \mathbb{P}(\text{pos. } 1-3) = {} & \ln \mathbb{P}(V \leftrightarrow V) \\
& + \ln \mathbb{P}(D \leftrightarrow -) \\
& + \ln \mathbb{P}(E \leftrightarrow E)
\end{aligned}
$$

side note on probability models

We usually project from:

- models of instantaneous rates of events – the "microscopic" scale (*sensu* Rivas & Eddy)

to:

- probabilities of descendant sequences – the "macroscopic" scale

Rivas and Eddy [preprint] have recently gone backwards

Rivas and Eddy L1 *vs* TKF91

TKF91 model infinitesimal time events

$$
\begin{array}{ll}
\underline{(s)}\,\mu\,\delta t & \text{Deletions of single ancestral residues.} \\
\underline{(i)}\,\mu\,\delta t & \text{Deletions of single inserted residues.} \\
\underline{(s+1+i)}\,\lambda\,\delta t & \text{Single-residue insertions.}
\end{array}
\tag{32}
$$

LI model infinitesimal time events

$$
\begin{array}{ll}
\underline{(s)}\,\mu_d\,\delta t & \text{Deletions of single ancestral residues.} \\
\underline{(i)}\,\mu_I\,\delta t & \text{Deletions of single inserted residues.} \\
\underline{(l+1+i)}\,\lambda_I\,\delta t & \text{Single-residue insertions.}
\end{array}
\tag{10}
$$
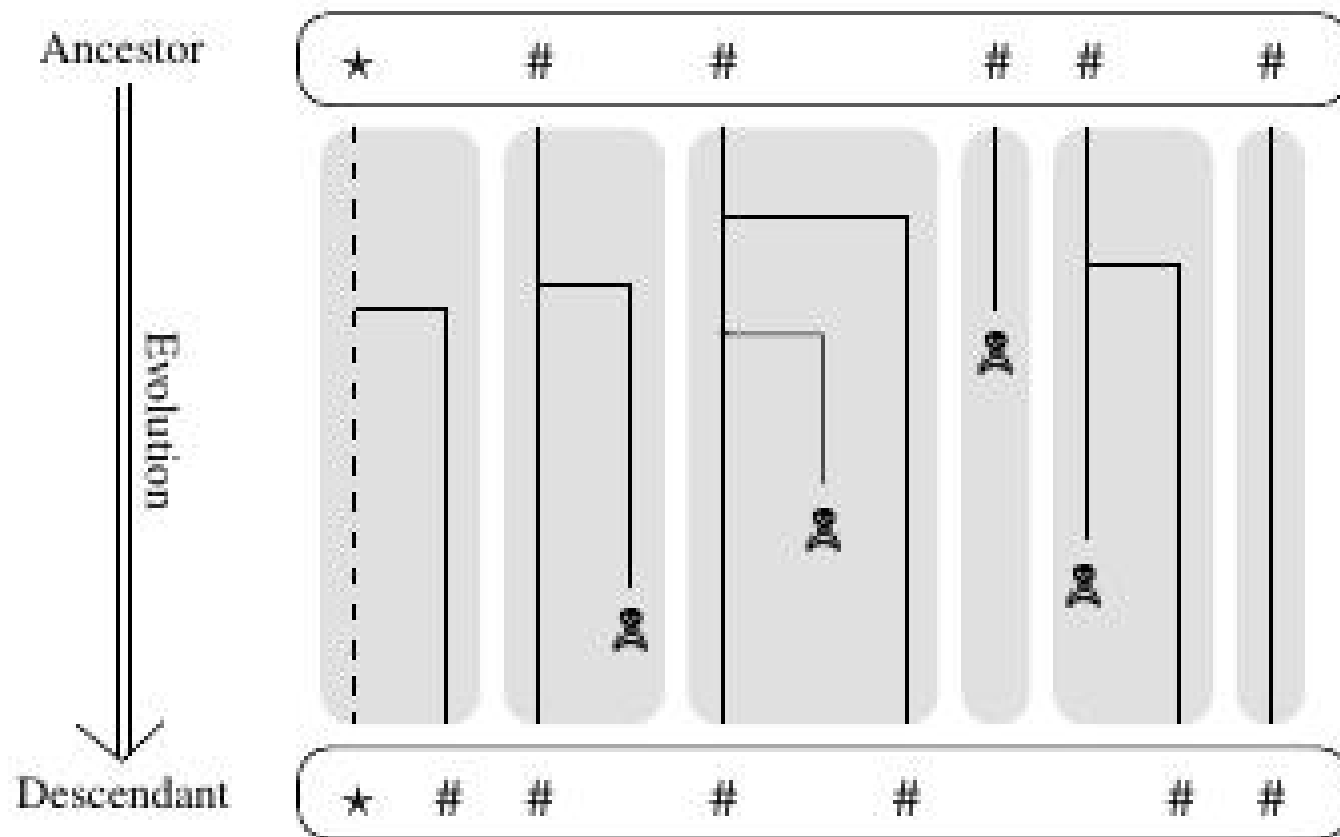
Fig 1.3 from a great review by Lunter *et al.* 2005

Rivas and Eddy models

Their models are natural to implement with dynamic programming

- linear reversible **LR** is a single-residue indel model similar to TKF91,
- Affine Insert Fragment **AIF** is an affine indel model similar to TKF92

Pairwise alignment summary

- The sum of the substitution and gap cost can serve as a proxy for the log-likelihood under a reasonable model (the **LR** or **AIF** model of Rivas and Eddy, preprint).

- Dynamic programming can let us find the alignment that has the highest likelihood.

from (Rausch and Reinert, 2011)

GAA-T
-AAC-
--ACT

Multiple sequence alignment is an ugly topic in bioinformatics

- Clever programming tricks help, but we still have to rely on *heuristics* – approaches that provide good solutions, but are not guaranteed to find the best solution.

- The additive scoring system suffers from the fact that we do not observe ancestral sequences.

- U is the set of unaligned sequences
- T is the genealogy tree that describes the ancestry of the sequences
- H is an indel history (specification of where all inserstions and deletions in the history of U occur on the tree T).
- A is an alignment of the sequences U

We might want:

- $\mathbb{P}(T \mid U)$ or $\mathbb{P}(A \mid U)$ or $\mathbb{P}(T, A \mid U)$. BaliPhy approximates these quantities, but it is tough to do for large datasets.

- $\hat{H}$ which maximizes $\mathbb{P}(H \mid U, T)$. ProtPal (Westesson et al., 2012) approximates this (but we have to know T)

Sum of Pairs scoring

Most MSA programs optimize a fairly strange score:

$$SP = \sum_i \sum_j w_{ij} d(A_i, A_j)$$

where A_i is the alignment pruned down to just sequence i, and w_{ij} is a weight for the comparison between sequences i and j.

$d(A_i, A_j)$ can be:

- a measure of the distance from A_i to A_j or
- a measure of the consistency of the alignment A with a pairwise alignment of i to j.

We usually cannot guarantee that we have found the alignment that optimizes the sum of pairs score.

Aligning more than two sequences
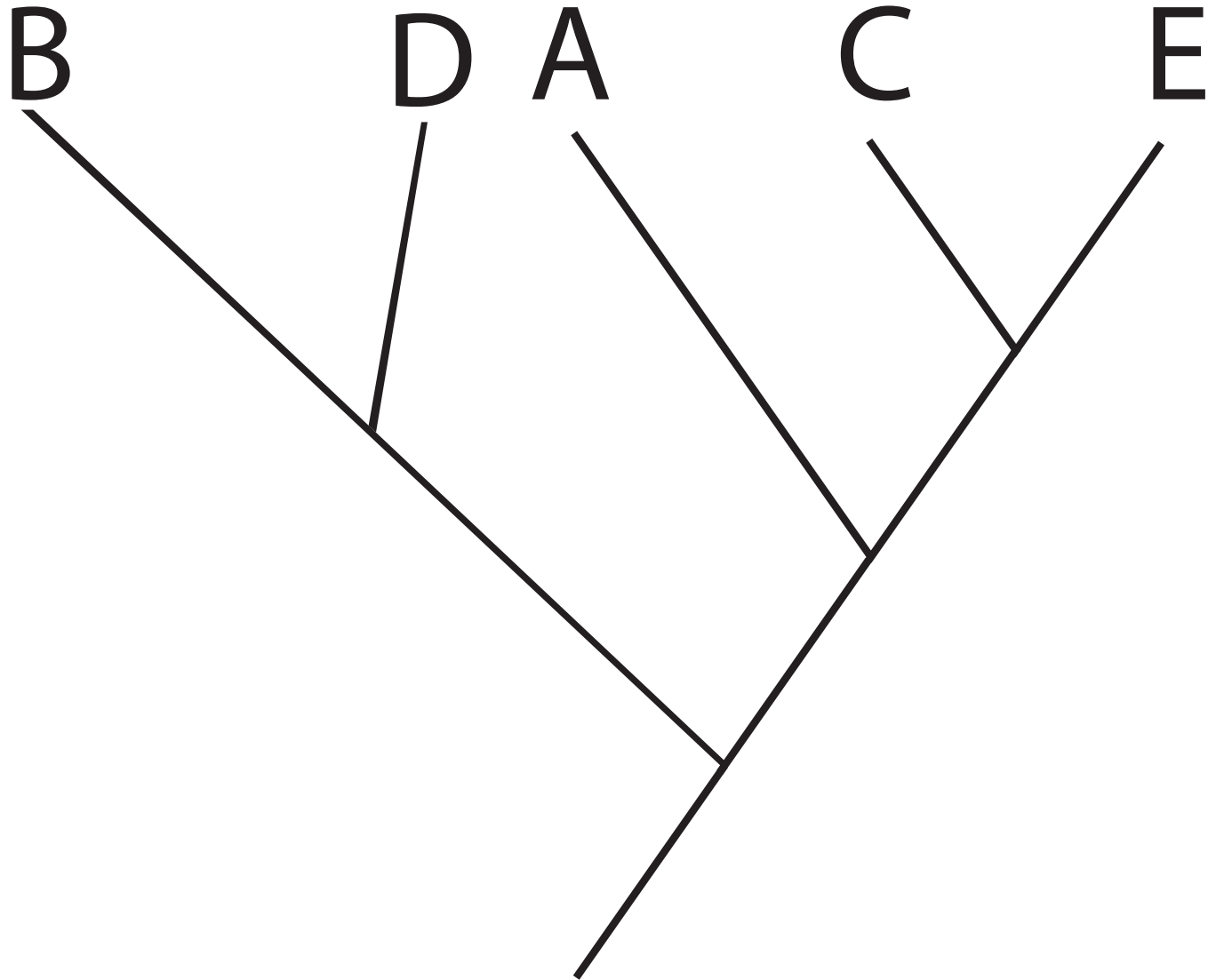
Progressive alignment

An approximate method for producing multiple sequence alignments using a guide tree.

- Perform pairwise alignments to produce a distance matrix

- Produce a guide tree from the distances

- Use the guide tree to specify the ordering used for aligning sequences, closest to furthest.

Feng and Doolittle 1987 and Higgins and Sharp, 1988

A   PEEKSAVTALWGKVNVDEVGG
B   GEEKAAVLALWDKVNEEEVGG
C   PADKTNVKAAWGKVGAHAGEYGA
D   AADKTNVKAAWSKVGGHAGEYGA
E   EHEWQLVLHVWAKVEADVAGHGQ

pairwise alignment

A   –
B   .17   –
C   .59   .60   –
D   .59   .59   .13   –
E   .77   .77   .75   .75   –

tree inference

A   PEEKSAVTALWGKVNVDEVGG
B   GEEKAAVLALWDKVNEEEVGG
C   PADKTNVKAAWGKVGAHAGEYGA
D   AADKTNVKAAWSKVGGHAGEYGA
E   EHEWQLVLHVWAKVEADVAGHGQ

+
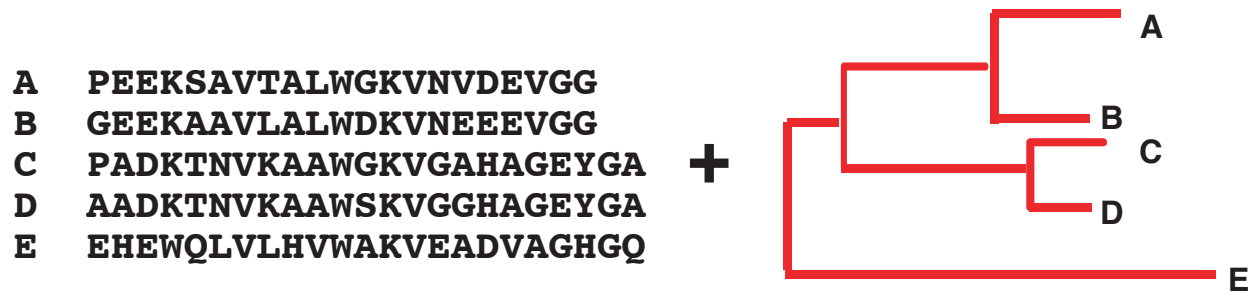
alignment stage

A   PEEKSAVTALWGKVN--VDEVGG
B   GEEKAAVLALWDKVN--EEEVGG
C   PADKTNVKAAWGKVGAHAGEYGA
D   AADKTNVKAAWSKVGGHAGEYGA
E   EHEWQLVLHVWAKVEADVAGHGQ

Alignment stage of progressive alignments

Sequences of clades become grouped as the algorithm descends the tree. Alignment at each step involves

- Sequence-Sequence,

- Sequence-Group, or

- Group-Group

Aligning multiple sequences

B   D A   C   E

0.1   0.2   0.15   .1

0.27

Seq-Seq

Seq-Seq

0.12

0.1   Seq-Group

0.09

Group-Group

Group-to-Group alignment
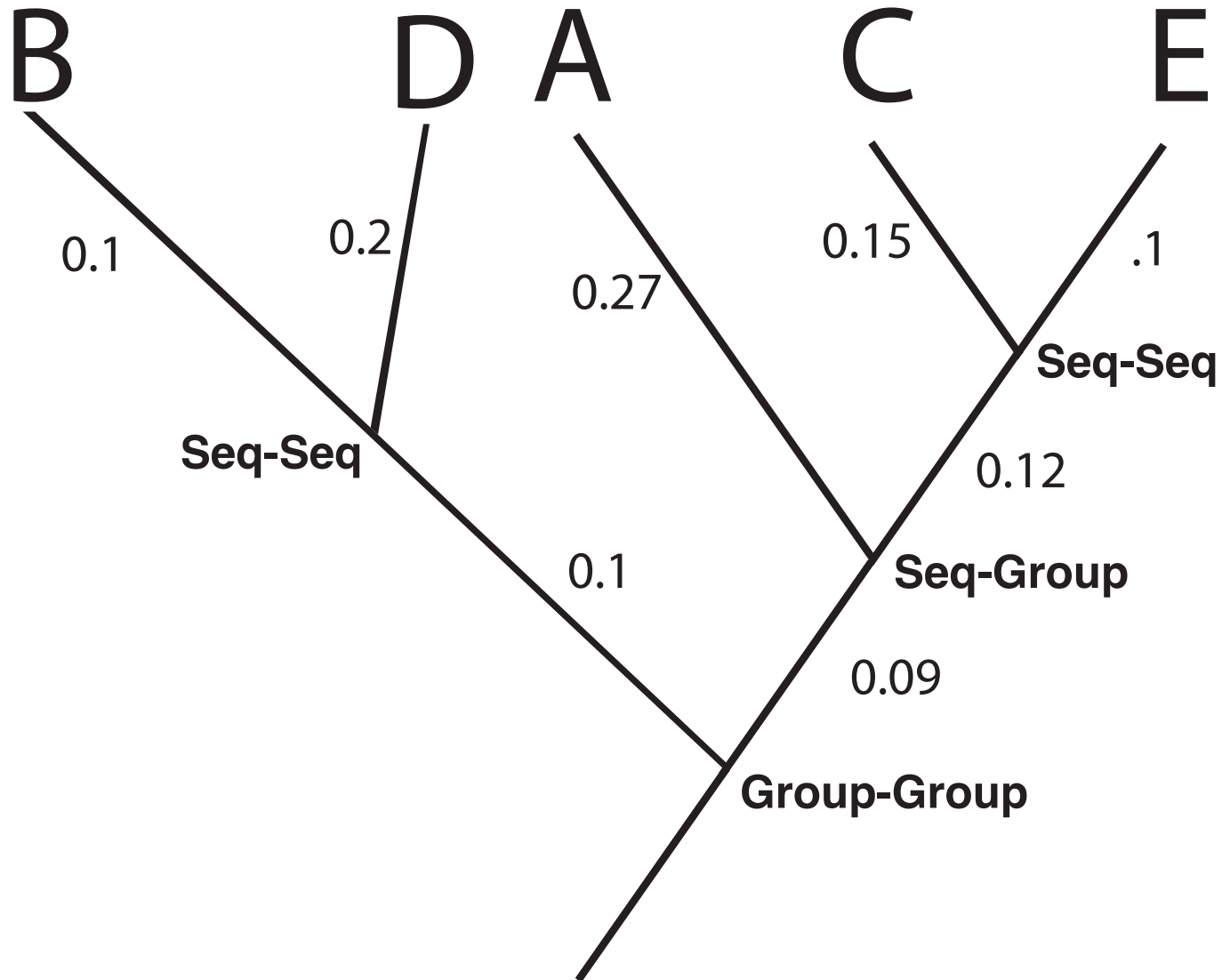
Group-to-group alignments

Adding a gap to a group means that every member of that group gets a gap at that position.
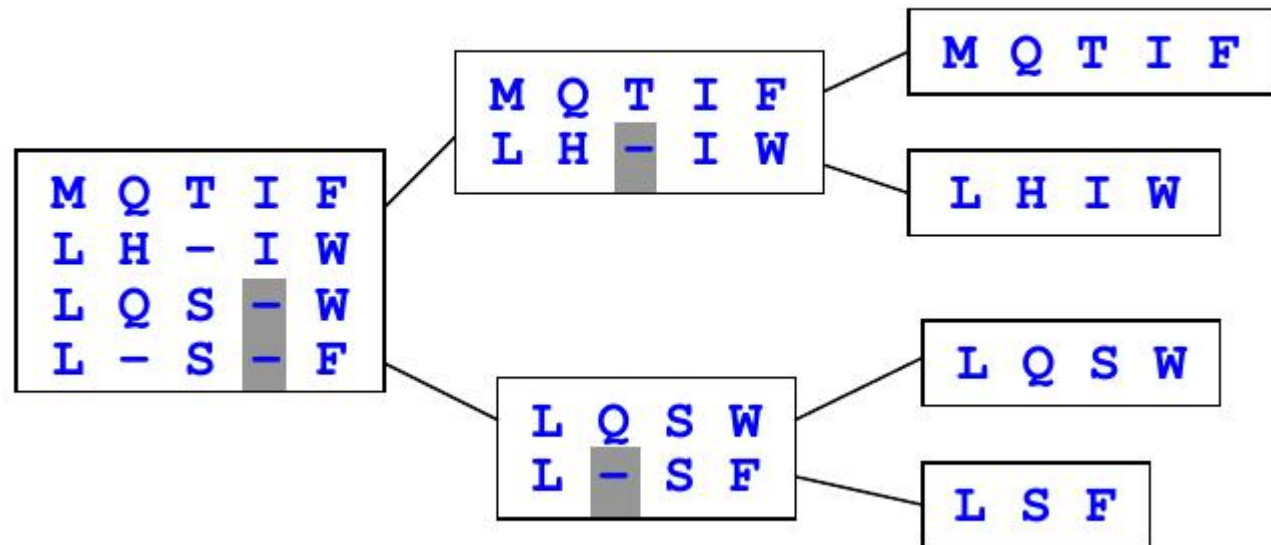
from Edgar (2004)

Group-to-group alignment

Usually the scores for each edge in the Needleman-Wunsch graph are calculated using a "sum of pairs" scoring system.

Many tools[1] uses weights assigned to each sequence in a group to down-weight closely related sequences so that they are not overrepresented - this is a weighted sum-of-pair scoring system.

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[1]e.g. Clustal and MAFFT

Greedy choices leading to failure to find the best alignment

Consider the scoring scheme:

match = 0   mismatch = -3   gap = -7

Guide Tree: Sequences:

Sp1   *Sp2*   *Sp3*
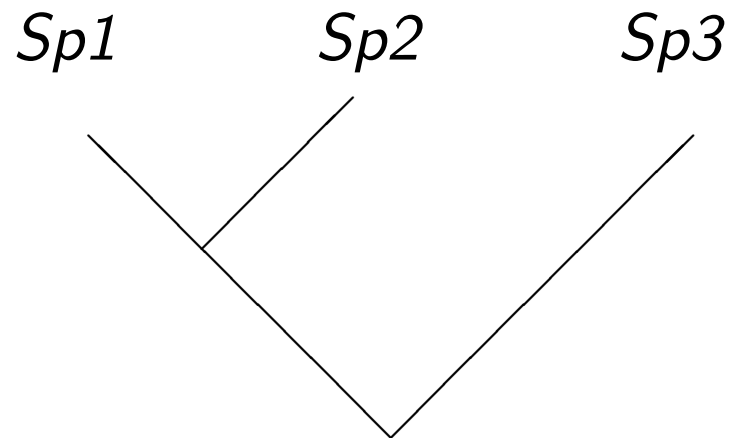
Sp1	GACCGTG
Sp2	GCCGTAG
Sp3	GACCGTAG

Greedy choices leading to failure to find the best alignment

match $= 0$ mismatch $= -3$ gap $= -7$

ungapped1*vs*2

Sp1	G	A	C	C	G	T	G		
Sp2	G	C	C	G	T	A	G		
Score	0	−3	0	−3	−3	−3	0	**Total=**	−12

would be preferred over gapped1*vs*2:

Sp1	G	A	C	C	G	T	−	G	
Sp2	G	−	C	C	G	T	A	G	
Score	0	−7	0	0	0	0	−7	0	**Total=**	−14

Adding a *Sp3* to ungapped1*vs*2:

Sp1	G	–	A	C	C	G	T	G
Sp2	G	–	C	C	G	T	A	G
Sp3	G	A	C	C	G	T	A	G

This implies 1 indel, and 4 substitutions. Score = -19 *

If we had been able to use gapped1*vs*2 then we could have:

Sp1	G	A	C	C	G	T	–	G
Sp2	G	–	C	C	G	T	A	G
Sp3	G	A	C	C	G	T	A	G

score = -14 *

* = "sort of…"

Score = -19 if we count events (but sum of pairs score would differ)
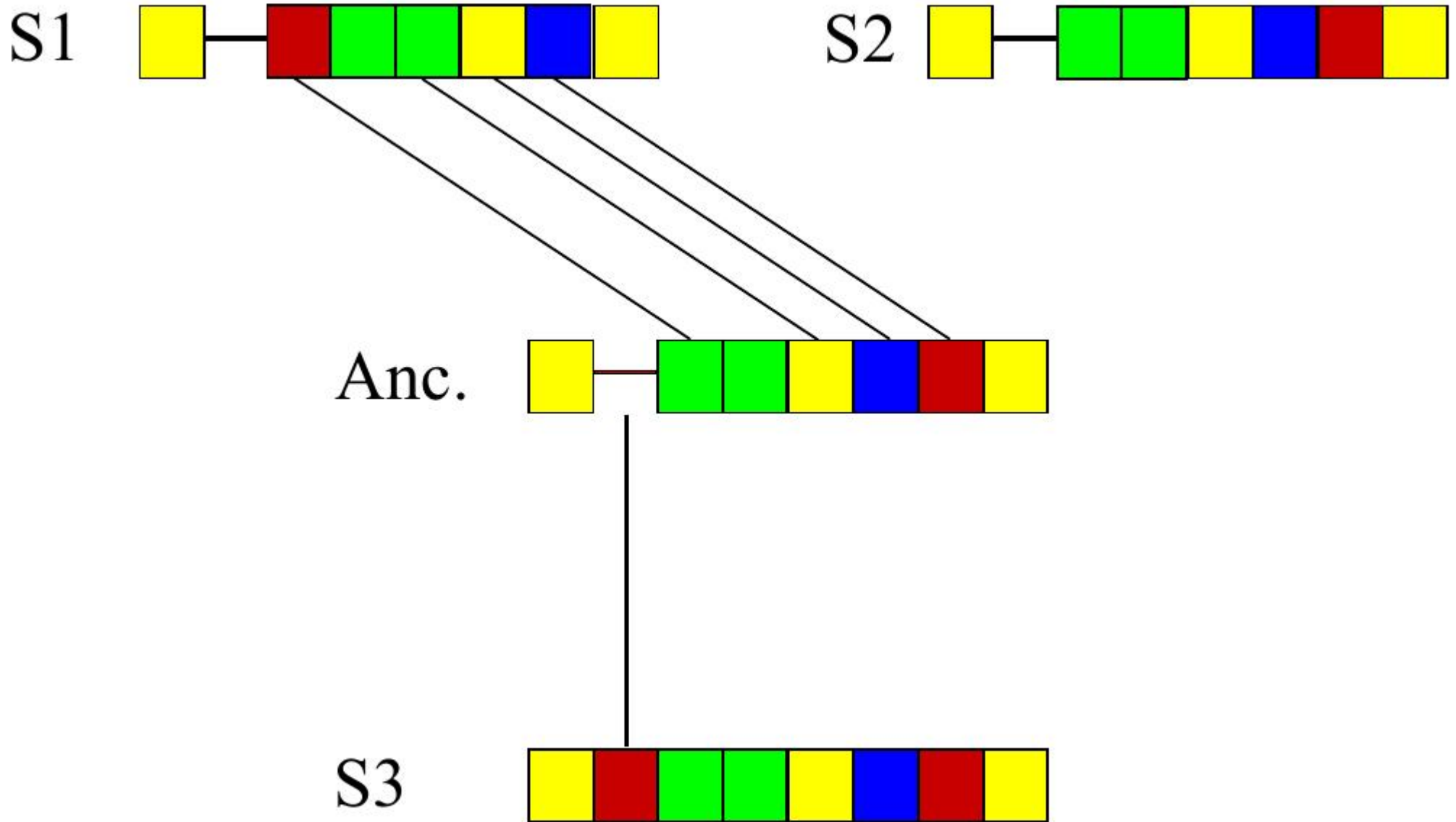
Score = -14 if we count events (but sum of pairs score would differ)
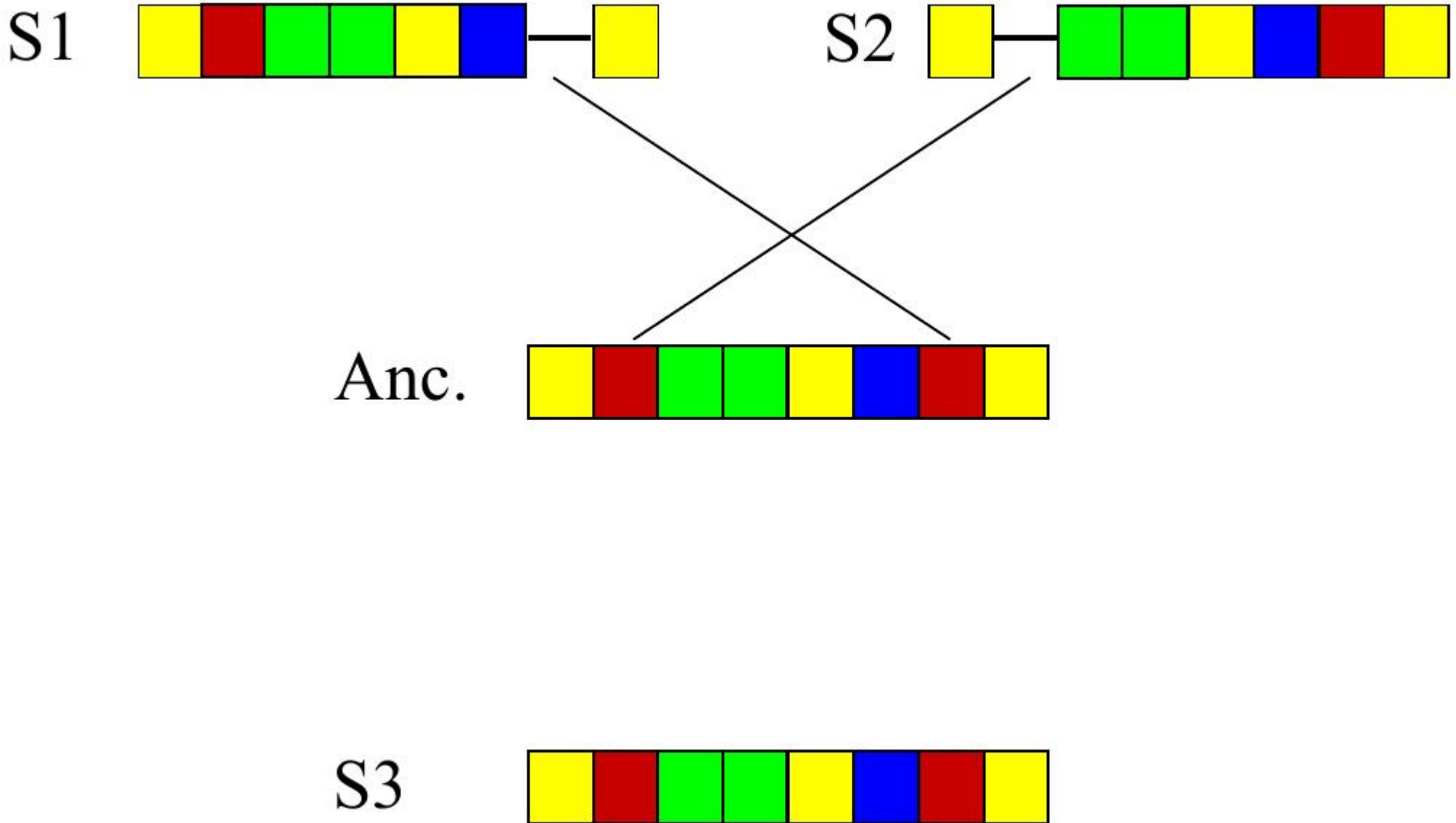
Polishing (aka "iterative alignment" can correct some errors caused by greedy heuristics)

1. break the alignment into 2 groups of sequences (often by breaking an edge in the merge tree).

2. realign those 2 groups to each other

3. keep the realignment if it improves the score

Opal also uses random 3-group polishing.

Weighted sum of pairs scoring system

Assigns a score for a group-to-group by averaging (or summing) the scores all of the implied pairwise alignments.

Frequently $w_{ij} = w_i w_j$

Group 1		
Seq	weight	AA
taxon A	0.3	V
taxon C	0.24	A
taxon E	0.19	I

Group 2		
Seq	weight	AA
taxon B	0.15	V
taxon D	0.25	M

$$D_{G1,G2} = \frac{\sum_i \sum_j w_i w_j d_{ij}}{n_i n_j}$$

	Group 1		
	Seq	weight	AA
taxon A	0.3	V
taxon C	0.24	A
taxon E	0.19	I

	Group 2		
	Seq	weight	AA
taxon B	0.15	V
taxon D	0.25	M

$$
\begin{aligned}
D_{G1,G2} &= \frac{\sum_i \sum_j w_i w_j d_{ij}}{n_i n_j} \\
&= \frac{1}{6}\left[d(V,V)w_A w_B + d(V,M)w_A w_D + d(A,V)w_C w_B \ldots \right. \\
&\quad \left. \ldots d(A,M)w_C w_D + d(I,V)w_E w_B + d(I,M)w_E w_D \right] \\
&= \frac{1}{6}(\mathbf{4} \times 0.3 \times 0.15 + \mathbf{1} \times 0.3 \times 0.25 + \mathbf{0} \times 0.24 \times 0.15 \ldots \\
&\quad \ldots -1 \times 0.24 \times 0.25 + \mathbf{3} \times 0.19 \times 0.15 + \mathbf{1} \times 0.19 \times 0.15) \\
&= 1.46225
\end{aligned}
$$

Sum-of-pairs is an odd scoring system

G A A A A L
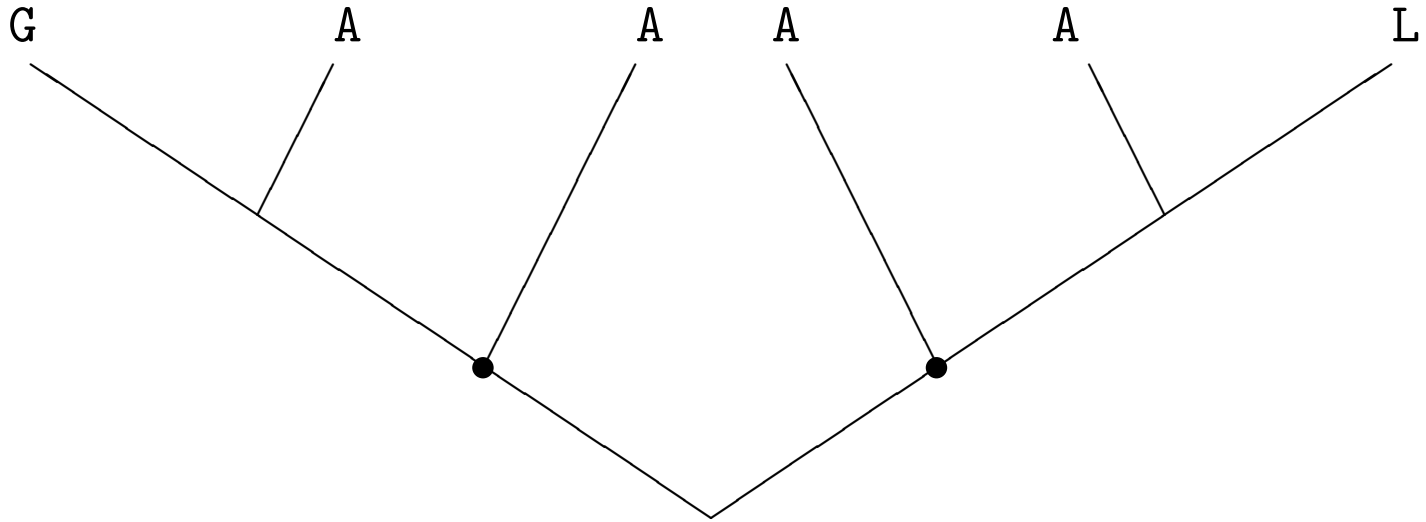
Imperfect scoring system. Consider one position in a group-to-group alignment:

```
(A,A,G)                (A,A,L)
```

$$(A,A,G) \leftrightarrow (A,A,L)$$
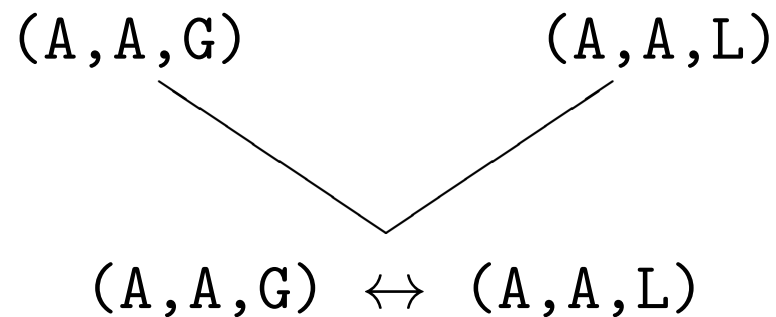
The sum-of-pairs score for aligning would be:

$$\frac{4}{9}(A \leftrightarrow A) + \frac{2}{9}(A \leftrightarrow L) + \frac{2}{9}(G \leftrightarrow A) + \frac{1}{9}(G \leftrightarrow L)$$

But in the context of the tree we might be pretty certain of an A↔A event

G   A   A   A   A   L

A ↔ A
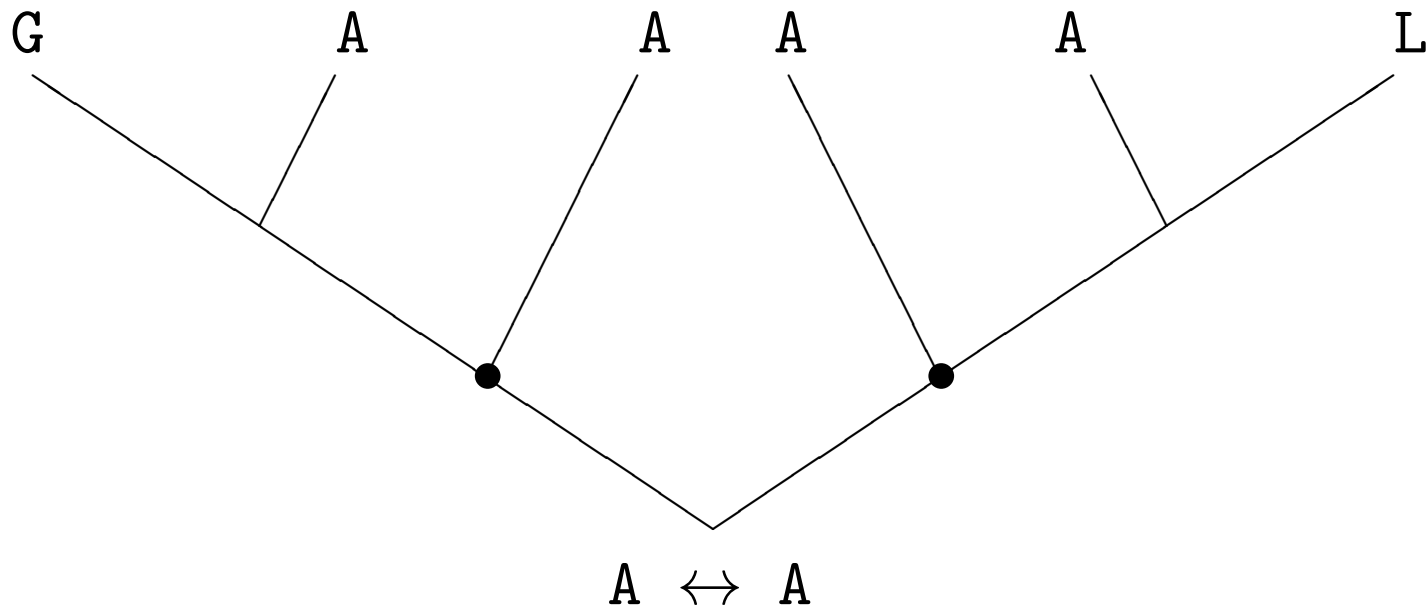
Note: weighted sum-of-pairs would help reflect the effect of ancestry better (but still not perfectly; sum-of-pairs techniques are simply not very sophisticated forms of ancestral sequence reconstruction).

"Consistency" based alignment

In our sum of pair scoring, we could just check how often each pair of sequences (one from G_h and one from G_v) display the same alignment that they did in pairwise alignment.

T-Coffee (Notredame et al., 2000) introduced the idea of performing group-to-group alignments during progressive alignment using this sense of consistency.

Indirect consistency arguments

In the pairwise alignments,

- *if* $h_{10} \sim g_{12}$
- *and* $p_{17} \sim g_{12}$
- *then* h_{10} should align with p_{17}

Probabilistic measures of "consistency"

The simplest assessment of the consistency of an MSA to pairwise alignments uses just the optimal pairwise alignment of each pair.

Opal (Wheeler and Kececioglu, 2007) uses some suboptimal alignments.

Do et al. (2005) made an important advance by proposing the use of the probability that two residues are aligned during consistency-based alignment (ProbCons).

The same sort of dynamic programming traversal of the alignment grid can give us a probability that 2 residues are aligned.

Sequence annealing

AMAP (Schwartz and Pachter, 2007) and FSA (Bradley et al., 2009) use a sequence annealing approach:

- start with trivial alignments (all residues opposite gaps),
- anchor regions by aligning long matches,
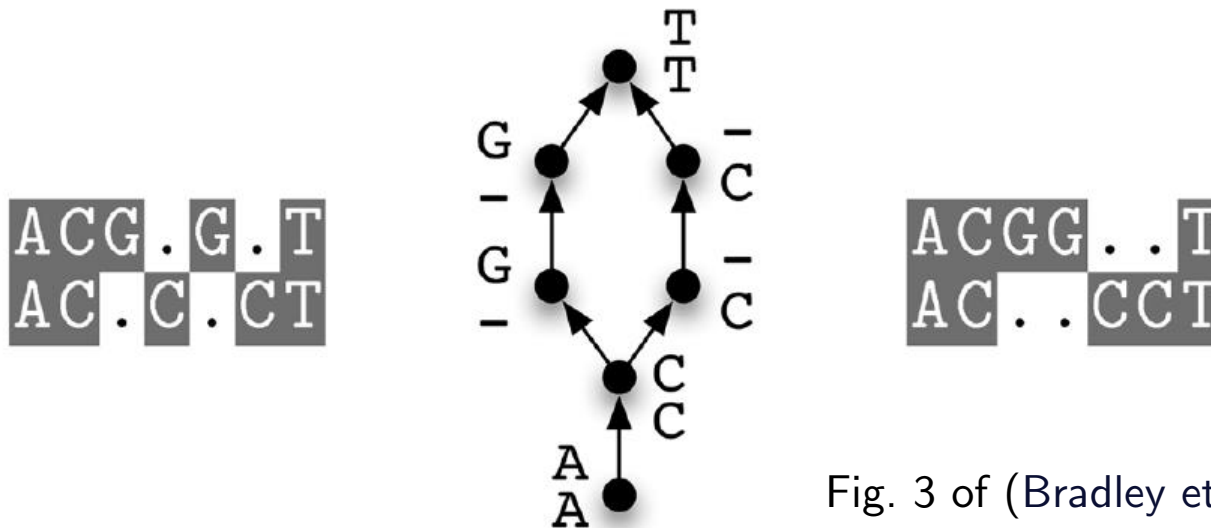- merge columns as long as the score keeps improving.

Fig. 3 of (Bradley et al., 2009)

Progressive alignment

- Uses a guide tree to change the MSA problem into a series of pairwise alignment problems;

- May not return the alignment with the best weighted sum-of-pairs scores. Early alignment decisions get "locked in" Most aligners try to polish the alignment, but we cannot guarantee that we have found the optimal alignment;

- Reconstruction of ancestral sequences is usually done in a quick-and-dirty, implicit fashion or is not done at all;
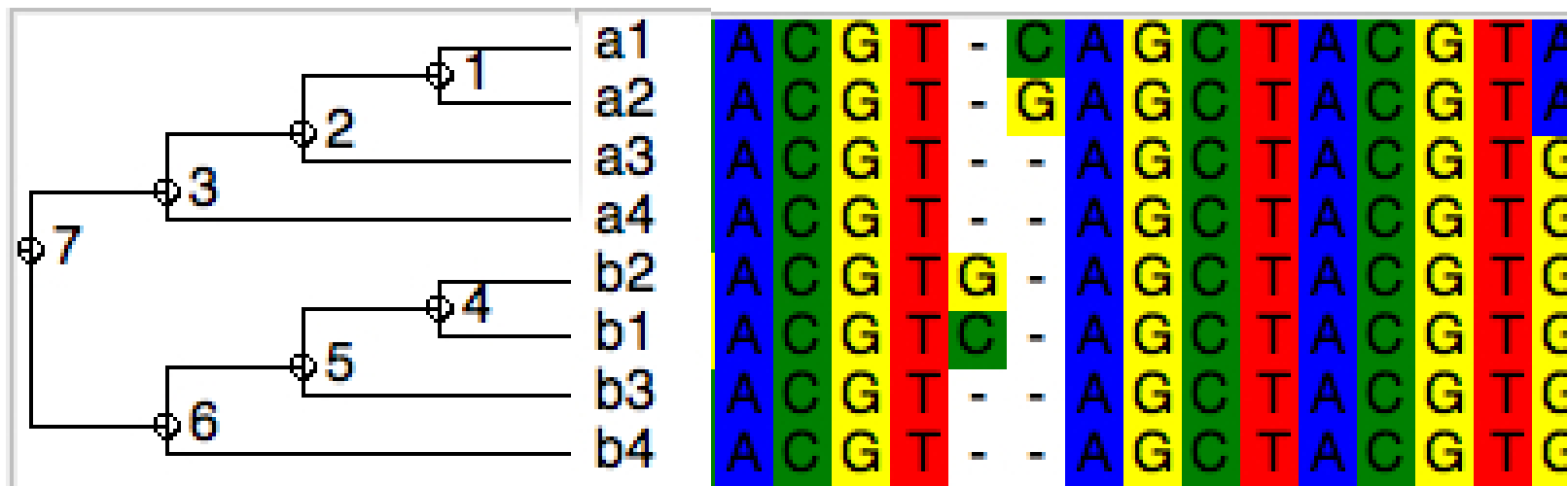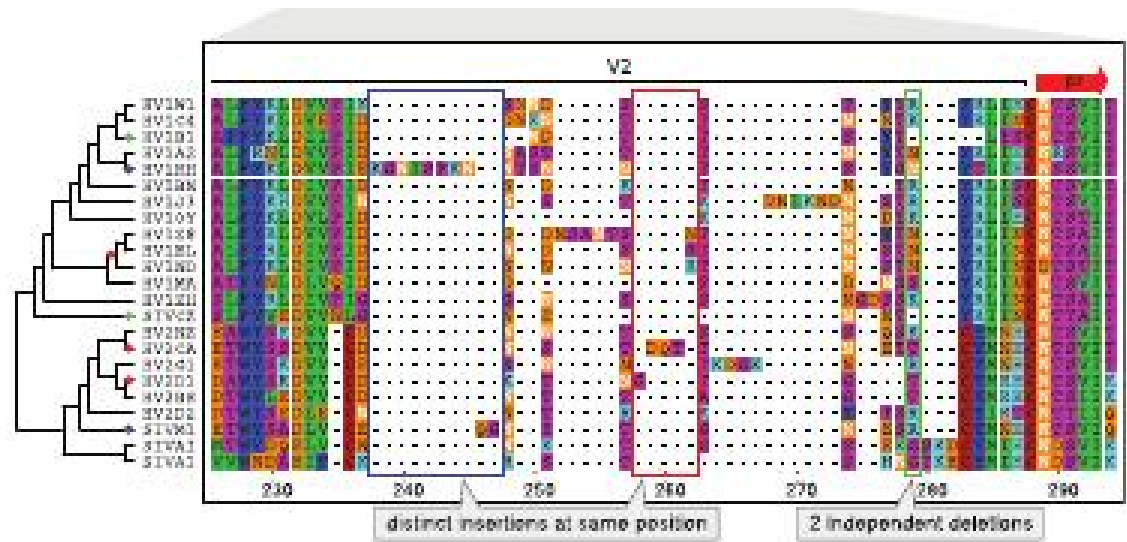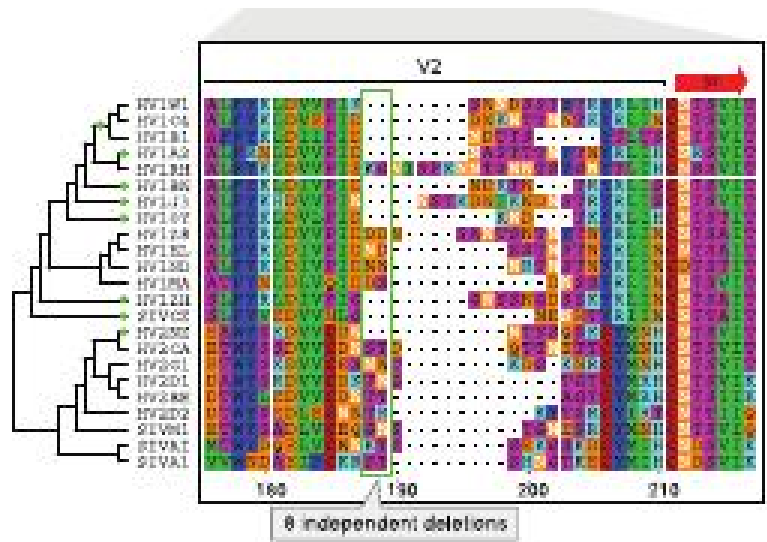
PRANK

Löytynoja and Goldman (2005) showed most progressive alignment techniques were particularly prone to compression because of poor ancestral reconstruction:
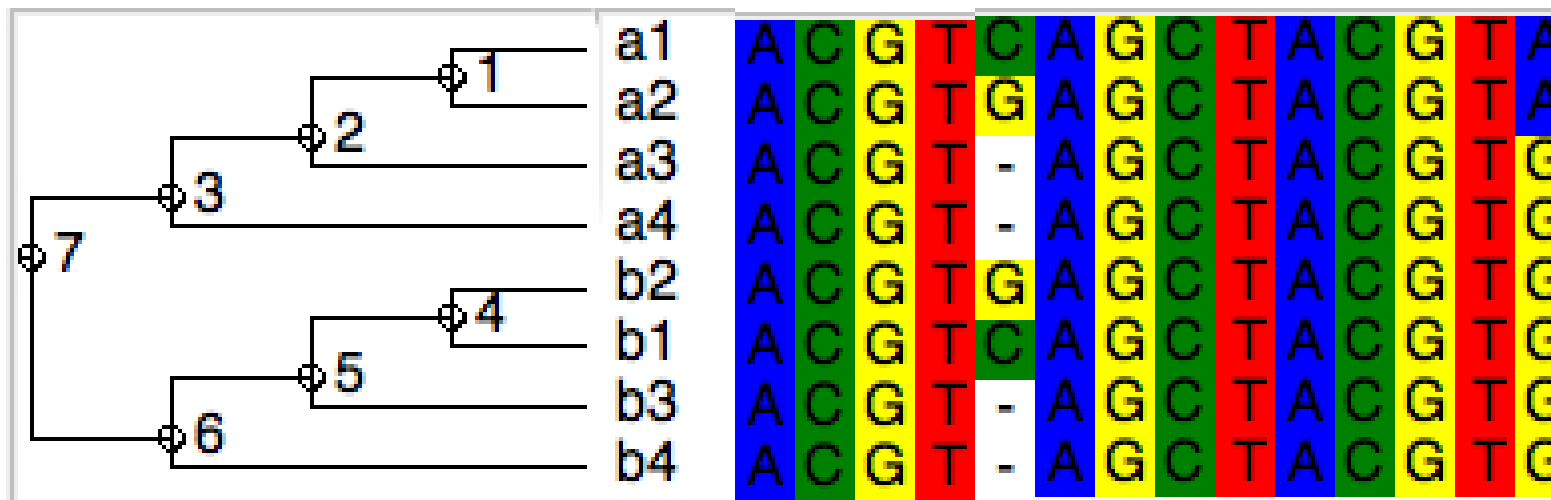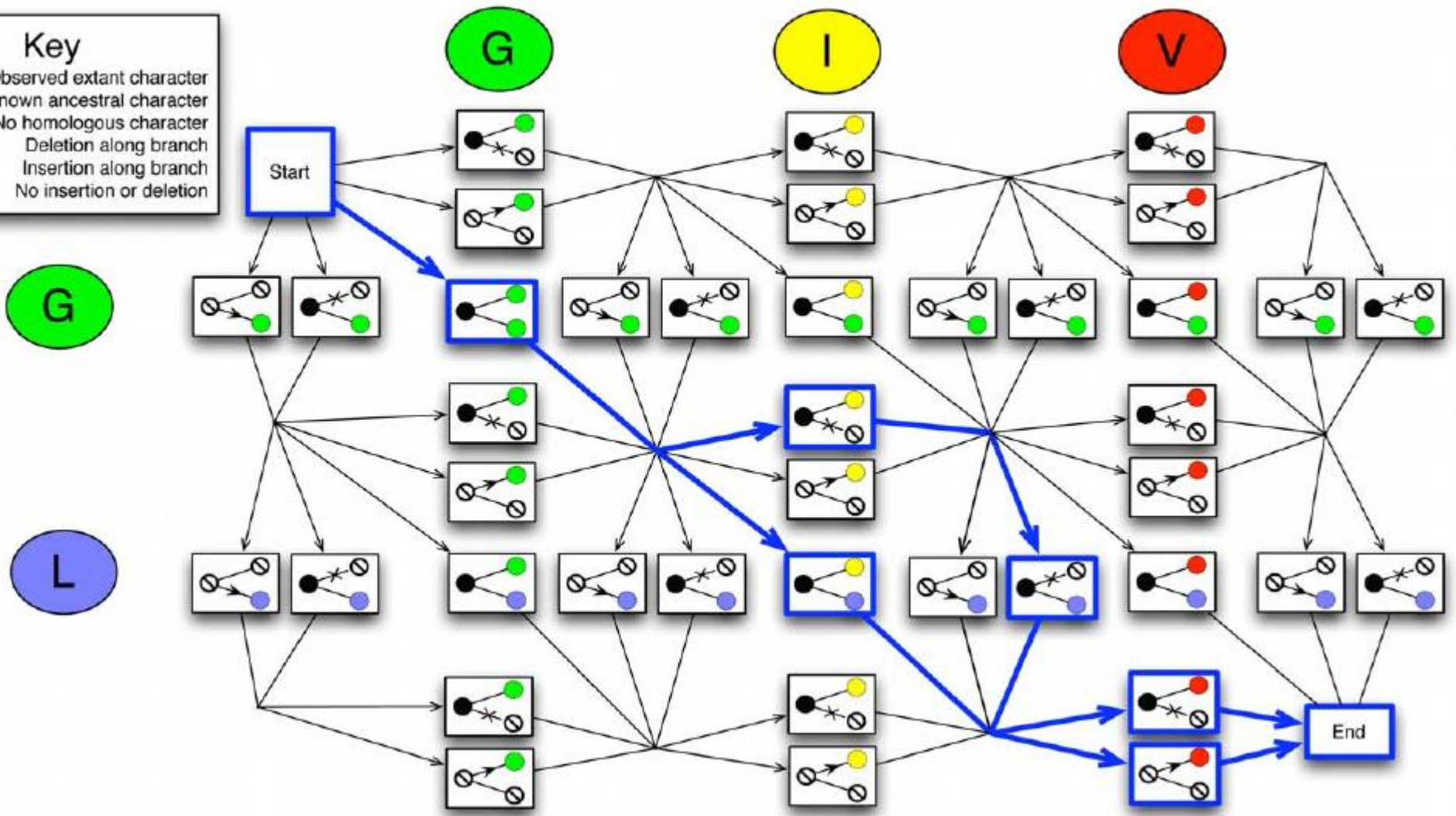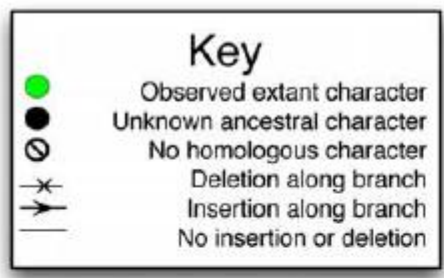
PRANK

Flagging inserted residues allows PRANK to effectively skip over these positions in the ancestor, producing more phylogenetically-sensible alignments:

ProtPal is similar to PRANK, but is retains a set of inferred ancestral seqs.
Fig. 5 of Westesson et al. (2012)

Impact of the guide tree

Using a guide tree can bias subsequent tree inference toward the guide tree.

This can also cause inflated support.

Ironically, this effect may be more of a problem for a more evolutionarily-sensible aligner such as PRANK or ProtPal!

Dealing with alignment ambiguity

(a)

	X			Y						Z		
	1	2	3	4	5	6	7	8	9	10	11	12
Outgroup	T	A	G	A	G	C	A	C	T	C	A	G
Taxon A	T	A	G	A	G	C	A	C	T	C	A	G
Taxon B	T	A	G	T	G	A	A	G	C	C	A	G
Taxon C	T	A	G	T	G	A	A	G	C	C	A	G
Taxon D	T	A	G		A	G	C			C	A	G
Taxon E	T	A	G		A	G	C			C	A	G

(b)

	X			Y						Z		
	1	2	3	4	5	6	7	8	9	10	11	12
Outgroup	T	A	G	A	G	C	A	C	T	C	A	G
Taxon A	T	A	G	A	G	C	A	C	T	C	A	G
Taxon B	T	A	G	T	G	A	A	G	C	C	A	G
Taxon C	T	A	G	T	G	A	A	G	C	C	A	G
Taxon D	T	A	G	A	G	C	–	–	–	C	A	G
Taxon E	T	A	G	A	G	C	–	–	–	C	A	G

(c)

	X			Y						Z		
	1	2	3	4	5	6	7	8	9	10	11	12
Outgroup	T	A	G	A	G	C	A	C	T	C	A	G
Taxon A	T	A	G	A	G	C	A	C	T	C	A	G
Taxon B	T	A	G	T	G	A	A	G	C	C	A	G
Taxon C	T	A	G	T	G	A	A	G	C	C	A	G
Taxon D	T	A	G	–	–	–	A	G	C	C	A	G
Taxon E	T	A	G	–	–	–	A	G	C	C	A	G

from M. S. Y. Lee, *TREE*, 2001

Dealing with alignment ambiguity - deletion

(a)

		X				Y					Z	
	1	2	3	4	5	6	7	8	9	10	11	12
Outgroup	T	A	G	A	G	C	A	C	T	C	A	G
Taxon A	T	A	G	A	G	C	A	C	T	C	A	G
Taxon B	T	A	G	T	G	A	A	G	C	C	A	G
Taxon C	T	A	G	T	G	A	A	G	C	C	A	G
Taxon D	T	A	G		A	G	C			C	A	G
Taxon E	T	A	G		A	G	C			C	A	G

		X		Z		
	1	2	3	10	11	12
Outgroup	T	A	G	C	A	G
Taxon A	T	A	G	C	A	G
Taxon B	T	A	G	C	A	G
Taxon C	T	A	G	C	A	G
Taxon D	T	A	G	C	A	G
Taxon E	T	A	G	C	A	G

|		X				Y					Z	
	1	2	3	4	5	6	7	8	9	10	11	12
Outgroup	T	A	G	A	G	C	A	C	T	C	A	G
Taxon A	T	A	G	A	G	C	A	C	T	C	A	G
Taxon B	T	A	G	T	G	A	A	G	C	C	A	G
Taxon C	T	A	G	T	G	A	A	G	C	C	A	G
Taxon D	T	A	G	?	?	?	-	-	-	C	A	G
Taxon E	T	A	G	?	?	?	-	-	-	C	A	G

Filtering

- GUIDANCE score (Penn et al., 2010) which reflects sensitivity of the alignment to poorly supported parts of the guide tree; and
- Head-or-Tails algorithm (Landan and Graur, 2007) which compares the alignment of the sequences to the alignment that you would get from reversed forms of the sequences.
- SOAP (Löytynoja and Milinkovitch, 2001) looks for parts of the alignment that are sensitive to alignment parameters
- trimAl (Capella-Gutiérrez et al., 2009) uses column properties and consistency
- TCS (Chang et al., 2014) uses consistency and provides column scores
- Gblocks, examine properties of the columns in the matrix.

Simultaneous tree inference and alignment

- Ideally we would jointly estimate with uncertainty

- Allows for application of statistical models to improve inference and assessments of reliability

- Just now becoming feasible: `BAliPhy` (Redelings and Suchard, 2005)
  See also: `POY` (Wheeler, Gladstein, Laet, 2002), `Handel` (Holmes and Bruno, 2001) , and `BEAST`(Lunter et al., 2005; Drummond and Rambaut, 2007).

- SATé (Liu et al., 2009) and PASTA (Mirarab et al., 2014) are iterative (back and forth between tree and alignment estimation)

SATé repeats the following steps until termination
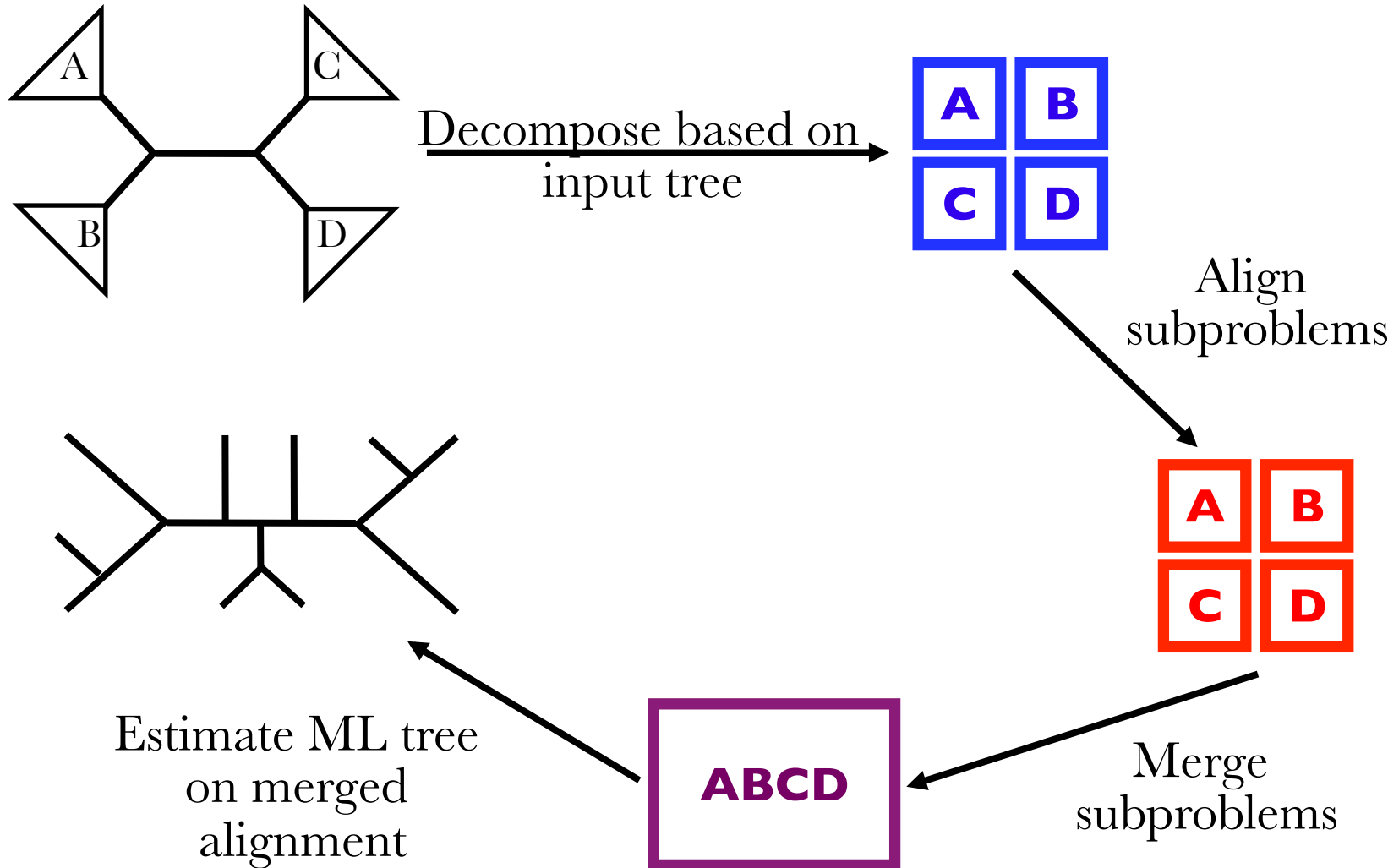
SATé simulation results
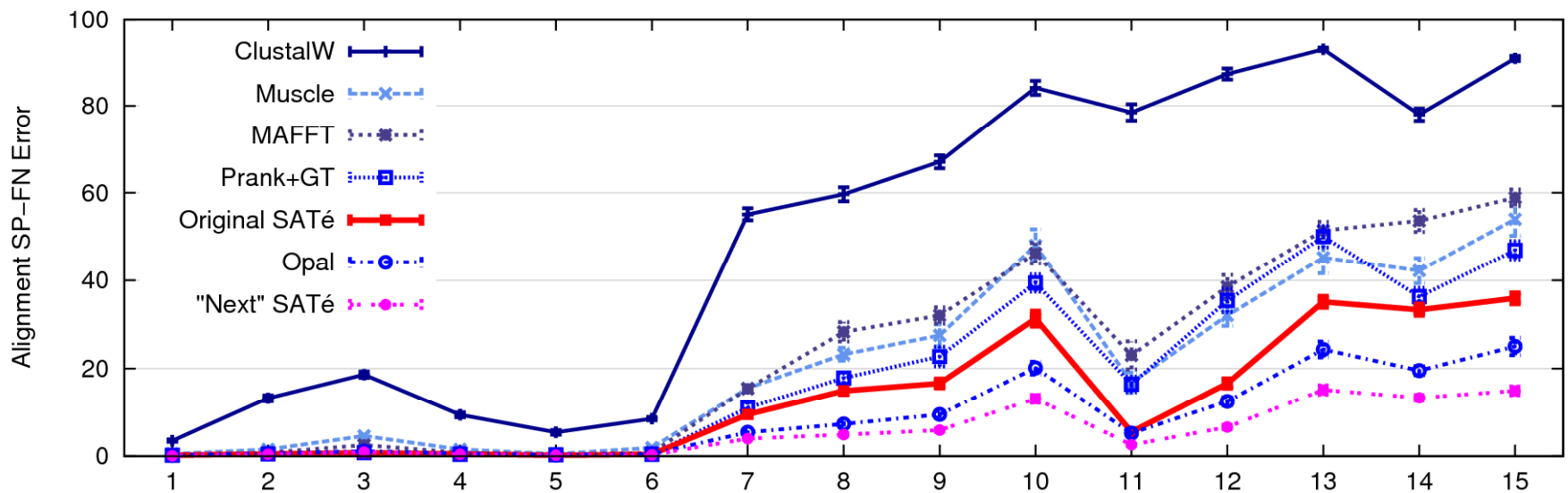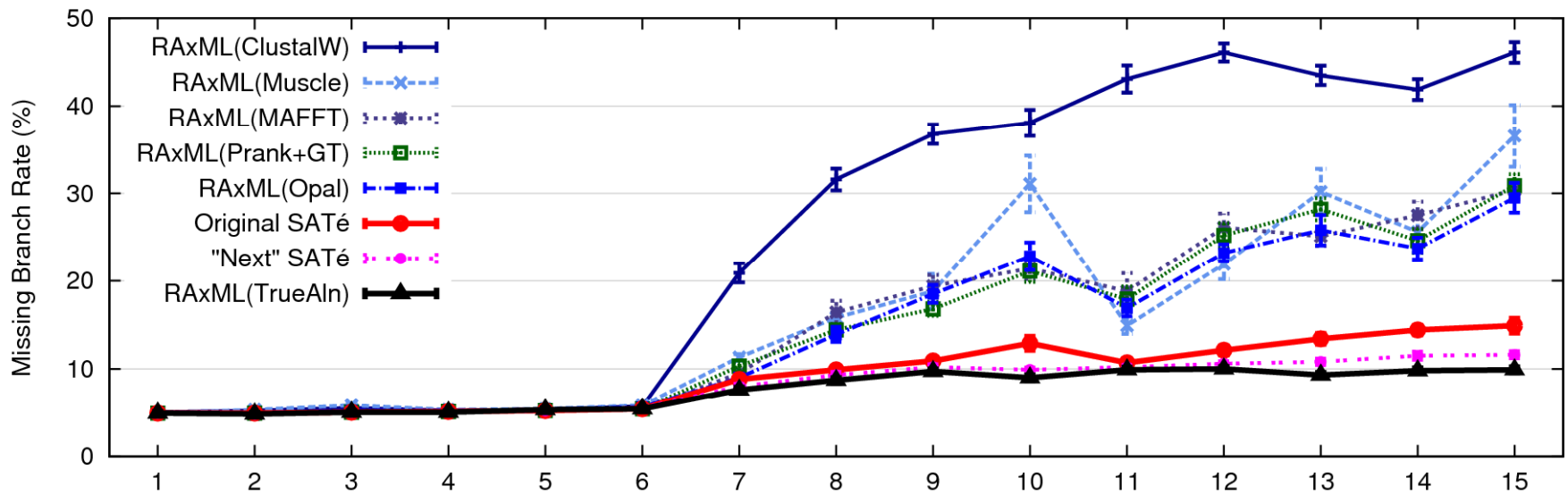
Conclusions

- Evolutionary multiple sequence alignment is still a very active area of research.
- We are hampered by:
  - lacking a good criterion to optimize (when T is unknown).
  - being forced to use rough heuristics to optimize the sum of pairs scores.
- Filtering throws away information, but may be helpful
- Most phylogenetic inference tools ignore information from the indel process (but see Rivas et al. (2008); Rivas and Eddy (2013) and GARLI's DIMM)

Recommendations

- See if BaliPhy can run on your data! If not, try FSA
- Look at your alignments
- Be aware that the standard aligners only consider substitutions and indels. If your data shows inversions, duplications, . . . there can be serious artifacts.
- if you have protein-coding sequences, use amino acid or codon-aware aligners
- if you have sequences from an RNA with important secondary structure, check out Infernal

A

FFT-NS-2 (Progressive method)
CPU time = 140 s
Wall-clock time = 35 s

B

L-INS-i (Iterative refinement method)
CPU time = 98 h
Wall-clock time = 6.5 h

C

Step 1: L-INS-i for full-length alignment
CPU time = 15 min
Wall-clock time = 1.0 min

Step 2: Profile alignments
CPU time = 2.5 min
Wall-clock time = 24 s

D

Full-length sequences
ITS2
5.8S rRNA
ITS1

Step 1: L-INS-i for full-length alignment
CPU time = 15 min
Wall-clock time = 1.0 min

Step 2: --6merpair --addfragments
CPU time = 1.5 min
Wall-clock time = 12 s

E

Step 1: L-INS-i for full-length alignment
CPU time = 15 min
Wall-clock time = 1.0 min

Step 2: --reorder --6merpair --addfragments
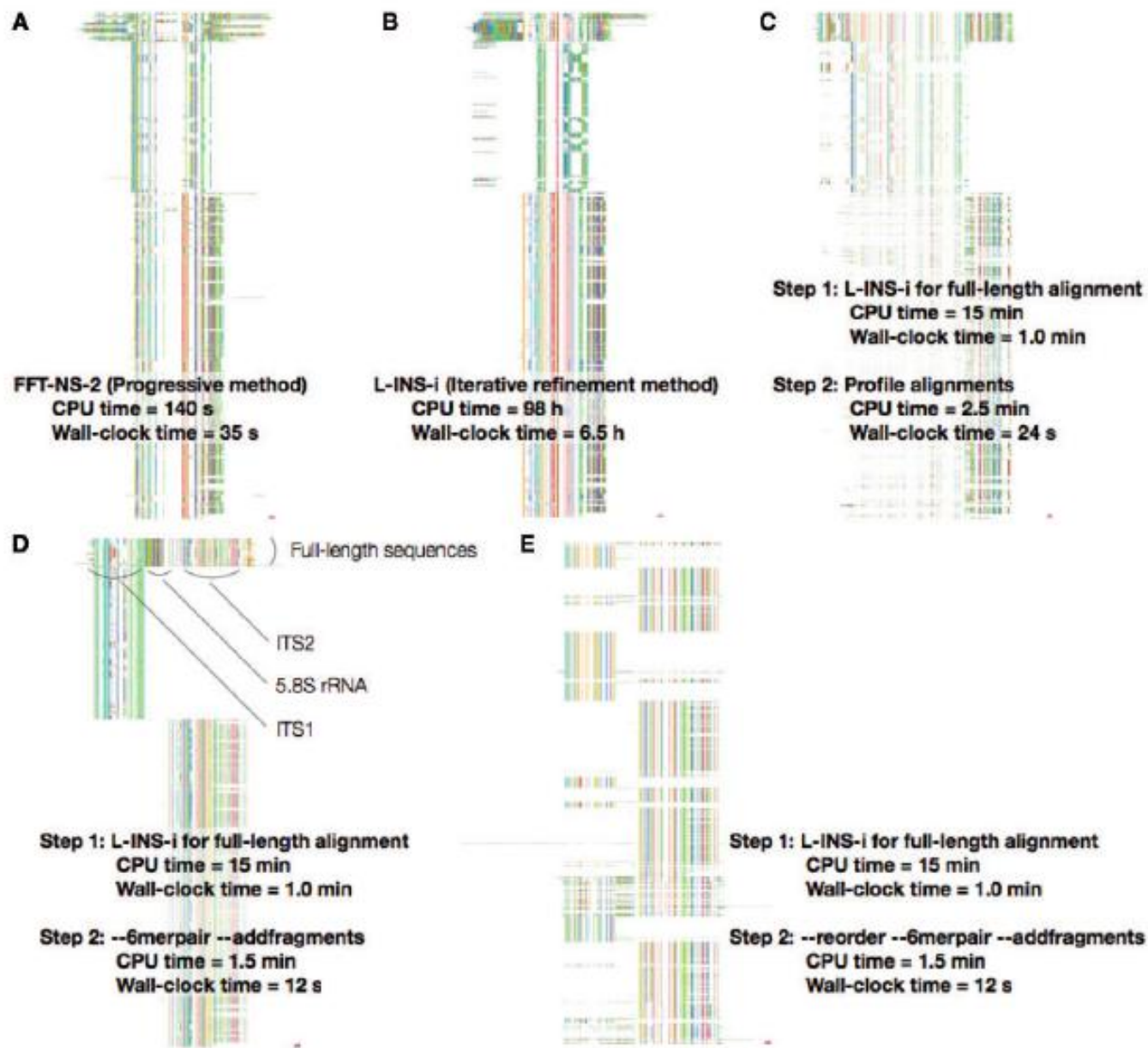CPU time = 1.5 min
Wall-clock time = 12 s

Fig. 2 from Katoh & Standley 2013

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Some of the tricks clustal uses to produce better alignments

- Chooses substitution matrix (PAM or BLOSUM series for amino acids) based on sequence similarity

- AA residues in a neighborhood affect gap opening penalty (easier to have gaps in hydrophilic loops)

- Gap penalties are raised if a column has no gaps, but there are gaps nearby.

- Low scoring alignments may be postponed until a later stage.

Terminal gaps

Using normal gap costs causes problems if one sequence is missing the starting or ending residues:

```
Pongo     VDEFKLIVEGELGRLFVVPTQ
Gorilla   VD-------GELGRLFVVPTQ
```

instead of :

```
Pongo     VDEFKLIVEGELGRLFVVPTQ
Gorilla   -------VDGELGRLFVVPTQ
```

(Methods that utilize local alignment information are more appropriate in these cases).

Using free terminal gaps to avoid this problem, but you have to watch out for:

```
Pongo     VDEPFRFKLTNRGTSHIILVAPR
Gorilla   ------VDEPNRGTSHIILVAPR
```